

Alere i Isothermal Amplification

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The old thinking. . . “it’s just the flu”

A bit of history

There are flu epidemics every 1 to 3 years for at least the last 400 years.

Pandemics (worldwide) occur around every 10 to 20 years.

History

Alere™

Hippocrates described flu back in the 5th century.

Columbus brought a devastating flu on his second voyage to the new world.

Spanish flu of 1918-1919 was the single greatest epidemic in history.

- 50 to 100 million people were killed (3-6% of the world's population!)
- Another 500 million were infected (1/3rd of the world's population)

Influenza Disease Burden

Estimated average global burden of seasonal influenza

- 600 million cases per year
- 3 million cases of severe illness
- 250,000 – 500,000 deaths

Concern is highest in

- The very young, the elderly
- persons with underlying health conditions
- pregnant women

Burden on Healthcare & Industry

Flu causes 30-50% increase in primary care consultations

2-3 fold increase in hospital admissions during epidemics

10% of all sickness absences from work

Impact in industrialised countries:

- US\$10-60 million / million population

Lost productivity:

- US\$12 billion / annum in US alone



Influenza A versus Influenza B

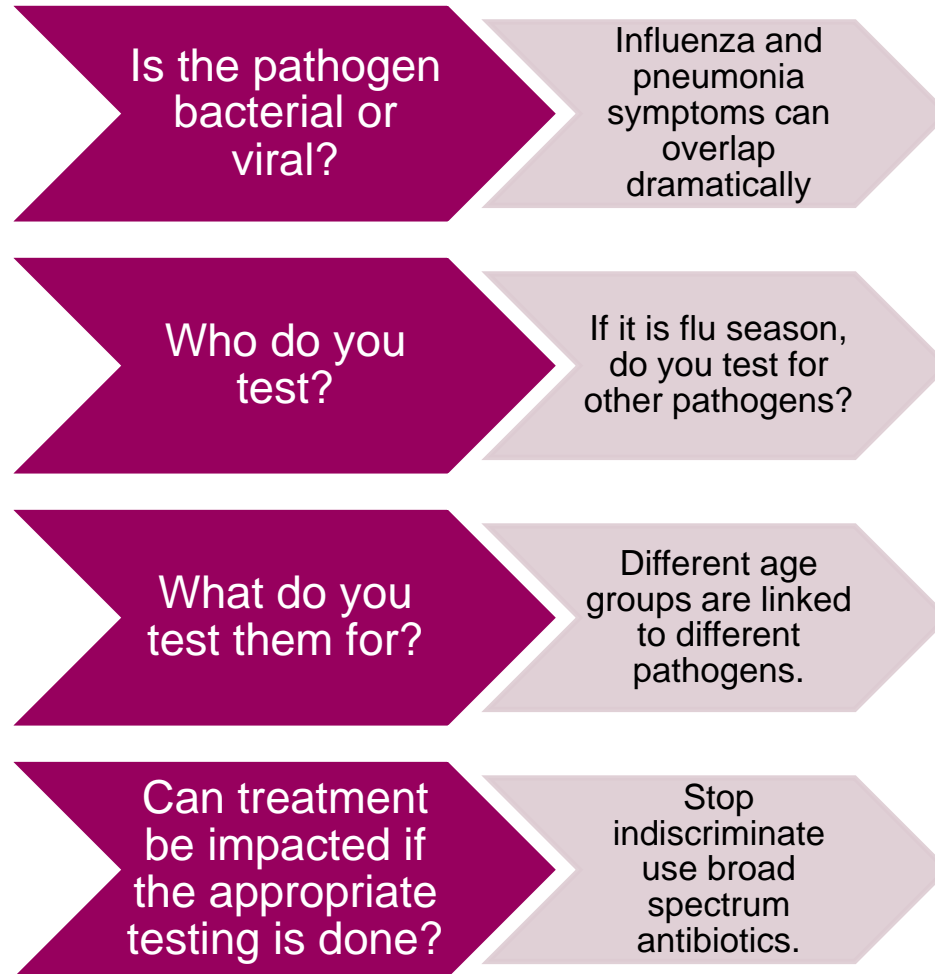
Influenza A

- More severe disease than B
- Can cause disease in a wide variety of animals

Influenza B

- Causes a milder flu, usually in the spring months

Treating Respiratory Diseases in the Emergency Department



Misuse of Antibiotics Can Lead to Other Medical Issues

Respiratory issues can be treated with fluoroquinolone

Disrupts normal intestinal flora

O27 strain of *C. difficile* is specifically resistant to fluoroquinolone

Diagnostic Methods for Influenza

Culture

DFA

PCR

Rapid Tests

Issues with Clinical Samples

Viral titer is highest in first 48 hours

Proper sample collection is necessary

Dilution in transport media

Rapid Tests

Pro

- Tests take minimal time
- Some tests are so simple that they can be CLIA-waived
- Can be used to triage patients
- Positive results can be used to rule out other issues like pneumonia so don't give unnecessary chest x-ray, antibiotics, etc.

Con

- Performance is not as good as culture, PCR, or DFA

Molecular Assays

Pro

- For respiratory specimens, high performance
- Same day results

Con

- Turn around time from lab may be extensive, especially if batching specimens
- Expensive
- May require experienced technicians, labs, dedicated equipment, etc.

The need is a test rapid enough to triage patients like a rapid test with molecular results.

What is Alere i?



What is Isothermal Amplification?

AlereTM i is a molecular diagnostic test using isothermal amplification

Isothermal amplification is not PCR but offers similar sensitivity and specificity

Uses a constant and low temperature so there is no need for thermal cycler

Very rapid amplification (many targets amplified in only 5-10 minutes)

Does not require DNA purification step, which adds time, complexity and cost

Ability to deploy an incredibly sensitive test for infectious diseases at the POC

Definition

NEAR, or Nicking Enzyme Amplification Reaction, enables the isothermal unwinding and subsequent amplification of very small amplicons using a set of target-specific templates (primers), a nicking endonuclease and a strand displacing DNA polymerase. Amplicon detection can be performed using a variety of probe formats including molecular beacons or by lateral flow sandwich.

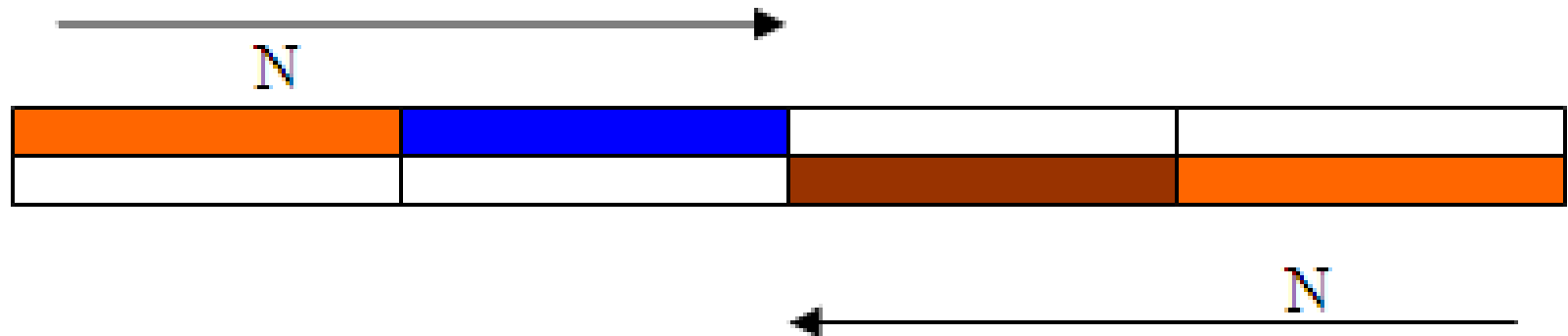
Initial Reaction

- Influenza is RNA virus
 - Perform reverse-transcriptase amplification
 - No purification of the nucleic acid

Amplification

The primed, duplex amplicon:

- blue recognizes a site on one strand of the target
- brown recognizes a nearly adjacent target on the other strand
- orange is an extra piece on each primer with a “nicking” site (N) and other “stabilizing” sequences
- there is a 10-12 bp “gap” between the “templates” (arrows); this enables use of a target-specific probe for product detection.
- after duplex synthesis, nicking at or near “N” enables strand displacement amplification, providing additional product that can be re-amplified using additional templates (primers).



iNAT Flu A/B Workflow



Consumables:



Sample Receiver



Test Base

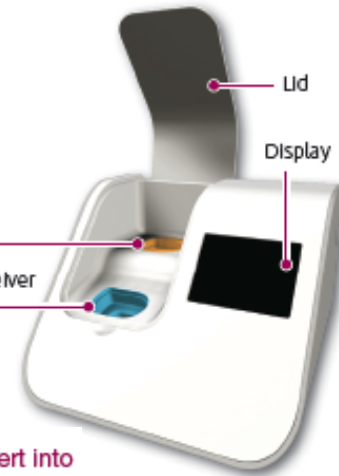


Transfer Cartridge



Cotton Swab

Device:



- 1 Touch "Run Test"**
This will begin the test process.
- 2 Enter and confirm patient data**
- 3 Open lid and insert Test Base into holder**
Test Base and holder are color coded.
- 4 Confirm test**
Touch "Confirm" to proceed.
- 5 Insert Sample Receiver into holder**
Sample Receiver and holder are color coded.
- 6 Wait 3 minutes for Sample Receiver to warm up**
Then remove foil from the Sample Receiver.
- 7 Take nasal swab from patient and insert into Sample Receiver**
- 8 Press Transfer Cartridge onto Sample Receiver**
Listen for click. Blue indicator on top of Transfer Cartridge will appear to confirm sample has been collected.
- 9 Lift and then attach Transfer Cartridge to Test Base**
Blue indicator on top of Transfer Cartridge will disappear to confirm sample has been dispensed.
- 10 Close lid and wait 10 minutes for test results**

Attach Test Base/Cartridge onto Sample Receiver and discard after completing test.

Influenza A/B: First Publications



CLINICAL EVALUATION OF THE ALERE™ i INFLUENZA A&B ASSAY, A POINT-OF-CARE RAPID NUCLEIC ACID AMPLIFICATION TEST FOR DETECTION OF INFLUENZA.

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ABSTRACT (Revised)

Background: Rapid testing and detection of influenza virus infection in an outpatient setting improves patient management. Although rapid antigen tests for influenza offer quick results, they lack adequate sensitivity. We report data from an ongoing clinical point-of-care (POC) evaluation using a new isothermal nucleic acid test (iNAT), the Alere™ i Influenza A&B assay. The assay is a rapid instrumented test for the qualitative detection of influenza A and B viral RNA from respiratory specimens. Results are available within 15 minutes from specimen collection, allowing for a quick turnaround time.

Methods: Specimens were collected during the 2012-2013 respiratory season. During the study 545 specimens were collected from study-eligible subjects (485 children and 60 adults). Two nasal swabs were collected from the same nostril of each subject. One swab was immediately eluted in viral transport media and viral cell culture was inoculated within 72 hrs and analyzed by fluorescent antibody test at 2 days and 5 days post inoculation. The other swab was tested directly with the Alere™ i Influenza A&B instrument within 2 hrs at room temperature following collection or up to 24 hours with refrigeration. Testing with the instrument was performed in a POC setting and completed according to step-by-step instructions provided on the instrument display. These results were compared to viral cell culture as the gold standard. A real-time influenza RT-PCR assay was used for discordant analysis.

Results: A total of 545 specimens were collected for testing. Of the specimens tested, two were excluded for uninterpretable viral culture results and one was excluded due to conflicting reference results. With testing of the Alere™ i Influenza A&B assay, there were 11 invalid results for influenza A and 15 for influenza B, resulting in invalid rates of 2% and 2.6%, respectively. For 531 valid influenza A and 527 valid influenza B results, the overall performance of the Alere™ i Influenza A&B assay compared to viral cell culture is summarized in Table 1. In children less than 18 years old, the sensitivity was 100% for detection of influenza A and 97.2% for influenza B. The specificities were 96.2% and 100% for influenza A and influenza B in children. For adults greater than or equal to 18 years old, the sensitivity and specificity for detection of influenza A were 100% and 96.6%, while for influenza B the sensitivity and specificity were each 100%.

Conclusions: The Alere™ i Influenza A&B assay was easy to perform and testing results were available within 15 minutes, rather than hours or days as compared to conventional molecular testing and viral cell culture. Overall, the Alere™ i Influenza A&B assay compared well to viral cell culture. The Alere™ i Influenza A&B assay combines the speed of a rapid antigen test with the sensitivity of a NAT in the POC setting and offers the opportunity to improve patient management.

INTRODUCTION

Influenza viruses cause respiratory illnesses that manifest clinically with common symptoms of fever, headache, body aches, extreme tiredness, nausea, stuffy nose or rhinitis, and dry cough (1). Most who have an influenza infection will have a mild illness, will generally not require medical care or antiviral drugs, and will recover in less than two weeks. Some people, however, are more likely to have influenza-related complications that result in being hospitalized and occasionally result in death. Persons at increased risk for severe disease or death include young children (especially those less than two years of age), older adults, and persons of any age with compromised respiratory, cardiac, or immune systems (2-3).

Rapid antigen tests are at the front line of diagnosis of many common infections including respiratory infections such as influenza virus. Though not as sensitive as molecular nucleic acid tests (NAT), these rapid antigen tests provide the quick turnaround time from sample collection to result necessary for patient management in point-of-care (POC) sites such as physician offices and busy emergency departments (4). The decreased sensitivity has long been a tradeoff for the acquisition of rapid results.

Isothermal NAT (iNAT) allow nucleic acid amplification in a very narrow temperature range, eliminating the need for expensive thermal cyclers and allowing for results to be obtained very quickly. The Alere™ i Influenza A&B assay is a rapid and automated *in vitro* diagnostic test for the qualitative detection of influenza A and B that uses iNAT technology to detect viral RNA from respiratory specimens. Results are available within 15 minutes, allowing for the possibility of turnaround time from sample collection to results reporting in less than half an hour. This technology provides a rapid, sensitive and specific molecular test for influenza virus that can be used in POC settings for timely diagnosis of influenza infections.

MATERIALS & METHODS

Specimens: A total of 545 respiratory nasal swab specimens were collected from study-eligible subjects (485 children and 60 adults) during 2012/2013 respiratory season at clinical sites in distinct regions around the United States. From each subject, two nasal swabs were collected from the same nostril, one for testing with shell vial viral culture and fluorescent antibody testing, and the other for direct testing on the Alere™ i Influenza A&B assay.

Alere™ i Influenza A&B Nucleic Acid Amplification Test: A schematic of the testing procedure is provided in Figure 1. Briefly, the test base is inserted into the appropriate color-coded receptacle, followed by placing the sample receiver into the corresponding color-coded receptacle. The sample receiver and buffer inside are then heated for three (3) minutes. Following the heating step, specimen is eluted from the swab directly into the sample receiver. The transfer cartridge is then used to transfer sample to the test base. Once this step has been confirmed by the user, the test proceeds and results are available within 10 minutes. All instructions are provided by animated graphics on the instrument display. For the study, positive and negative quality control swabs were processed every day before testing of study specimens. Nasal swabs were tested with the Alere™ i Influenza A&B assay within 2 hours at room temperature following collection or up to 24 hours with refrigeration. All specimens were tested by directly eluting the nasal swab specimen into the buffer in the sample receiver.

Respiratory Viral Culture: Viral culture was performed using shell vials followed by analysis with fluorescent antibodies.

Discordant Analysis: Prodesse ProFlu+ real-time RT-PCR was performed on discordant specimens.

Specimen Analysis: A total of 545 specimens were enrolled into the study. Two specimens yielded uninterpretable viral culture results. One specimen yielded conflicting reference results. With Alere™ i Influenza A&B testing, there were 11 specimens with invalid influenza A results and 15 specimens with invalid influenza B results.

Figure 1. Overview of the Alere™ i Influenza A&B Assay



RESULTS

Table 1. Detection of Flu A and B by Alere™ i Influenza A&B in comparison with viral culture.

Alere™ i Influenza A&B	Influenza Type	TP	FP	TN	FN	Total	% Sensitivity (95% CI)	% Specificity (95% CI)	PPV (95% CI)	NPV (95% CI)
v. Culture	Flu A	89	49	377	1	517	98.2 (93.2-99.9)	98.7 (93.9-99.9)	98.2 (93.3-99.9)	99.7 (93.3-99.9)
	Flu B	87	17	437	6	527	97.2 (92.4-99.6)	99.3 (93.9-97.7)	97.9 (93.3-99.4)	99.8 (93.3-99.4)

RESULTS (cont.)

Table 2. Detection of Flu A and B by Alere™ i Influenza A&B in comparison with viral culture after discordant analysis.

Alere™ i Influenza A&B	Influenza Type	TP	FP	TN	FN	Total	% Sensitivity (95% CI)	% Specificity (95% CI)	PPV (95% CI)	NPV (95% CI)
v. Culture after PCR discordant analysis	Flu A	118	8	370	0	516	100 (98.2-100)	97.9 (94.9-99.8)	100 (97.7-100)	100 (98.7-100)
	Flu B	84	0	441	2	527	97.7 (91.1-99.6)	100 (93.3-100)	100 (94.8-100)	98.6 (93.3-99.8)

Table 3. Detection of Flu A and B in children and adults by Alere™ i Influenza A&B in comparison with viral culture after discordant analysis.

Specimen Detection	TP	FP	TN	FN	Total	% Sensitivity (95% CI)	% Specificity (95% CI)	PPV (95% CI)	NPV (95% CI)	
Flu A	Children	59	8	324	0	401	100 (98.2-100)	98.2 (95.1-99.9)	100 (97.7-100)	100 (98.4-100)
	Adults	17	0	87	0	104	100 (77.1-100)	97.3 (93.4-99.4)	100 (95.5-100)	100 (92.1-100)
Flu B	Children	78	0	378	2	458	97.2 (93.4-99.5)	100 (93.7-100)	100 (93.2-100)	100 (97.9-100)
	Adults	14	0	80	0	94	100 (73.2-100)	100 (73.2-100)	100 (73.2-100)	100 (92.7-100)

CONCLUSIONS

- In comparison to viral culture as the gold standard prior to discordant analysis, overall sensitivity of the Alere™ i Influenza A&B POC test was 98.2% for the detection of influenza A, and 97.2% for the detection of influenza B from direct nasal swabs. The specificities were 96.2% and 96.3% for influenza A and B, respectively.
- After discordant analysis with Prodesse ProFlu+ real-time RT-PCR, the overall sensitivity of the Alere™ i Influenza A&B assay was 100% for the detection of influenza A, and 97.2% for the detection of influenza B from direct nasal swabs. The specificities were 97.9% and 100% for influenza A and B, respectively.
- In children, when compared to viral culture (after discordant analysis by Prodesse ProFlu+ real-time RT-PCR) overall sensitivity of the Alere™ i Influenza A&B assay for the detection of influenza A was 100%, and 97.2% for detecting influenza B from direct nasal swabs. Specificity was 96.2% for influenza A and 100% for influenza B in children. In adults, overall sensitivity of the Alere™ i Influenza A&B assay for the detection of both influenza A and influenza B was 100%. Specificity was 96.6% for influenza A and 100% for influenza B.
- The Alere™ i Influenza A&B assay is a simple and easy to use molecular diagnostic test with no off-board lysis step. The test has the ability to generate results within 15 minutes from the time of specimen collection. Importantly, the test demonstrates impressive sensitivity for the detection of both influenza A and B in clinical respiratory specimens obtained from both children and adults.


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Influenza A/B: First Publications




UMKC School of Medicine

EVALUATION OF THE RAPID ALERE™ i INFLUENZA A&B NUCLEIC ACID AMPLIFICATION TEST USING RESPIRATORY SPECIMENS COLLECTED IN VIRAL TRANSPORT MEDIUM

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Children's Mercy Hospitals & Clinics
Kansas City

ABSTRACT

Background: Molecular nucleic acid-based laboratory tests are among the most sensitive tests available for diagnosis of viral infections. The Alere™ i Influenza A&B assay is a rapid isothermal nucleic acid amplification test for the sensitive detection of Influenza A and B in respiratory specimens. Results are available within 15 minutes.

Methods: A total of 236 salvaged frozen respiratory specimens collected from August 2006 through June 2012 were included in the study. The respiratory specimens were nasopharyngeal aspirates (NPA) or nasopharyngeal wash/aspirates (NPA) collected in 3 ml universal transport medium (UTM) from children. The results of the Alere™ i Influenza A&B assay were compared to R-ibo™ shell vial culture and ProFlu™ RT-PCR assay. All testing was completed within 24 hours of thawing the specimens. The Alere™ i Influenza A&B assay was performed by adding 200 µl of respiratory specimen directly to the instrument for lysis and isothermal amplification. Viral cultures were performed using R-ibo™ shell vial and stained with influenza A&B case antibody on day 2. Nucleic acids were extracted with the ProFlu™ Nuc B DNA/RNA assay kit and frozen at -20°C prior to batch-testing by real-time RT-PCR assay.

Results: The performance of the Alere™ i Influenza A&B assay compared to viral culture and PCR testing is summarized in Table 1. A total of 105 influenza A and 50 influenza B were detected by culture and 110 influenza A and 25 influenza B were detected by real-time RT-PCR. The Alere™ i Influenza A&B assay detected 105 influenza A and 50 influenza B specimens in comparison with culture. The influenza A test results of the Alere™ i Influenza A&B assay were 90 true positive (TP), 7 false positive (FP), 121 true negative (TN), and 7 false negative (FN). When compared to PCR, the influenza A results were 103 TP, 1 FN, 11 TN, and 0 false positive (FP) (1.7%).

Conclusions: The test performance of Alere™ i Influenza A&B assay with respiratory specimens collected in UTM compared favorably with that results from viral culture versus real-time RT-PCR. The Alere™ i Influenza A&B assay test results were available in minutes versus hours for conventional PCR and days ahead of the culture results. The Alere™ i Influenza A&B assay test is simple and easy to perform without the need for specialized nucleic acid extraction. The Alere™ i Influenza A&B assay provides both the speed of a rapid antigen test with the sensitivity of a nucleic acid based molecular test.

MATERIALS & METHODS

Specimens: A total of 236 salvaged frozen respiratory specimens collected in 3 ml of UTM from August 2006 through June 2012 at Children's Mercy Hospital and Clinics in Kansas City, Missouri, USA, were included in the study. There were 150 NRG and 70 NPA from children ages 10 months to 16 years that were included. Each specimen was de-identified and given a unique, site-specific study identifier prior to enrollment into the study.

Alere™ i Influenza A&B Nucleic Acid Amplification Test: A schematic of the testing procedure is provided in Figure 1. Briefly, the test base is inserted into the appropriate color-coded receptacle, followed by placing the sample receiver into the compartment color-coded receptacle. The sample receiver and buffer is then heated for three (3) minutes. Following the heating step, specimen is added directly to the sample receiver. The transfer cartridge is then used to transfer sample to the test base. Once this step has been confirmed by the user, the test procedure and results are available within 10 minutes. All instructions are provided by an integrated graphic on the instrument display. For the study, positive and negative quality control assays were processed every day before testing of study specimens. Specimens were tested with the Alere™ i Influenza A&B assay within 24 hours of thawing. All specimens were tested by adding 200 µl of thawed specimen in UTM directly to the buffer in the sample receiver. The testing procedure described above was then followed.

Respiratory Viral Culture: Viral culture was performed using R-ibo™ shell vial from Diagnostic Hybrids. The viral cultures were stained with Light Diagnostics™ SemiFluor™ FluA/FluB dual antibody from DIB Millipore on day 2 of culture.

RT-PCR: RT-PCR was performed with the ProFlu™ Nuc B Assay for detection of Influenza A, Influenza B, and RSV/Gem-ProdC. The assay kit performs on all thawed specimens according to the manufacturer's instructions on the Cepheid SmartCycler II Real Time Instrument with the software version 1.7a. Nucleic acids were extracted in batches daily and stored frozen at -20°C until real-time RT-PCR testing. Total nucleic acid was extracted from the specimens along with positive and negative extraction controls with the NucB DNA/RNA automated extractor from bioMérieux.

RESULTS

Table 1. Comparison of Alere™ i Influenza A&B Assay versus Respiratory Viral Culture and ProFlu™ RT-PCR

Alere™ i Influenza A&B	Specimen	TP	FP	FN	TN	Total	Sensitivity (%)		Specificity (%)		PPV (%)		NPV (%)	
							95% CI	95% CI	95% CI	95% CI	95% CI	95% CI		
v. Culture	Influenza A	90	7	121	7	235	93.3	94.5	100	100	100	94.5	100	99.9
	Influenza B	50	-	174	-	224	100	100	100	100	100	100	100	100
v. PCR	Influenza A	103	1	115	12	231	98.2	99.2	100	100	100	99.1	100	99.9
	Influenza B	50	-	174	-	224	100	100	100	100	100	100	100	100

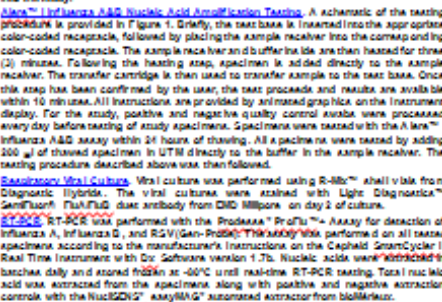


Figure 1. Overview of the Alere™ i Influenza A&B Assay

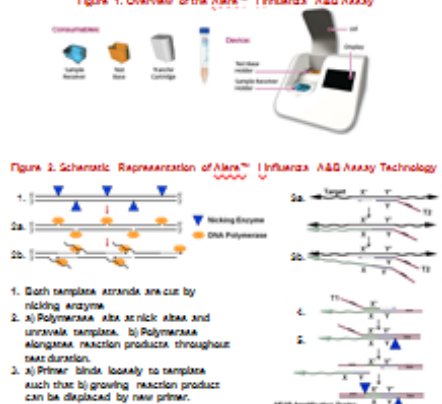


Figure 2. Schematic Representation of Alere™ i Influenza A&B Assay Technology

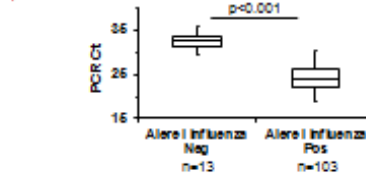


Figure 3. Comparison of PCR Ct values of Alere™ i Influenza A&B positive and negative specimens.

CONCLUSION

- Using viral culture as the gold standard, overall sensitivity of the Alere™ i Influenza A&B assay was 93.3% for the detection of influenza A, and 100% for detection of influenza B. The specificities were 94.5% and 100% for influenza A and B, respectively.
- In comparison to PCR, overall sensitivity of the Alere™ i Influenza A&B assay for the detection of influenza A was 98.2%, and 100% for detecting influenza B. Specificity was 99.2% for influenza A and 100% for influenza B.
- The PCR cutoff for the detection of Influenza A and B viruses for specimens collected in 3 ml of UTM based on the ProFlu™ Nuc B Assay was determined to be a Ct of 30. For the i Influenza Neg specimens the Ct range was 29.4 - 35.6 (1st Quartile 29.4 - 34.2; 3rd Quartile 30.7 - 35.2). For the i Influenza Pos specimen the Ct range was 18.1 - 30.4 (1st Quartile 18.1 - 22.1; 3rd Quartile 24.1 - 25.3).
- The Alere™ i Influenza A&B assay is a simple and easy to use molecular diagnostic test with a 15-minute turnaround time. The assay has the ability to generate results within 15 minutes from the time of testing the test.

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Acknowledgements

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Influenza A/B: First Publications

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Jeremiah Bell, Aista Bonner, Daniel M. Cohen, Robert Birkhahn, Ram Yogen, Wayne Trimer, Jason Cohen, Elizabeth Palavecino, and Rangaraj Selvarangan

ABSTRACT (Revised)

MATERIALS & METHODS

RESULTS (cont.)

Direct NS vs Culture

Flu A: 100% / 97.9%

Flu B: 97.7% / 100%

REFERENCES

Acknowledgements

EVALUATION OF THE RAPID ALERE™ I INFLUENZA A&B NUCLEIC ACID AMPLIFICATION TEST USING RESPIRATORY SPECIMENS COLLECTED IN VIRAL TRANSPORT MEDIUM

Jeremiah Bell and Rangaraj Selvarangan

ABSTRACT

MATERIALS & METHODS

RESULTS

VTM vs ProFlu+ PCR

Flu A: 88.8% / 98.3%

Flu B: 100% / 100%

REFERENCES

Acknowledgements

Conclusions:

- ✓ Compared well to viral cell culture
- ✓ Demonstrates impressive sensitivity
- ✓ Combines speed of a RAT with sensitivity of a NAT in POC setting
- ✓ Offers the opportunity to improve patient management

Advantages of Rapid Flu Tests

AlereTM

Accurate determination of who needs medication

- Antiviral medication AND antibiotics
- Avoid exacerbation of bacterial & viral resistance
- In pandemic, can help qualify who gets medication

Early treatment of high risk patients, to

- Reduce complications
- Reduce spread
- Reduce healthcare burden

Cost benefits

- Directed therapy
- Reduced morbidity and mortality reduces hospital costs

